

Review on Retention of Placenta in Dairy Cows and its Economic and Reproductive Impacts

Tolera Tagesu Tucho

Jimma University, College Of Agriculture and Veterinary Medicine, School of Veterinary Medicine
Jimma, Ethiopia

Abstract

Fetal membranes or what is known as “placenta” is an essential organ for prenatal transfer of nutrients and oxygen from the dam to the fetus. The normal separation of fetal membranes consists of complex hormonal process that starts before parturition in cows, which drops within short time after birth. It normally drops within short time post partum. If the placenta is not expelled within certain time (12 h post calving and 3 h post foaling), it is defined as being retained placenta (RP). It's a condition where all or part of the placenta or membranes are left behind in the uterus during the third stage of labour. Retained placenta creates a number of problems following pulling of microorganisms into the uterus causing its inflammation, decreased milk yield, longer calving intervals, reduction of fertility, longer calving interval, and reduce conception rate. RP causes great economic losses, mainly due to decreased milk yield and loss due to infertility. The number of risk factors of retained placenta includes stillbirth, dystocia, abortion, twin, fetotomy, induction of parturition, cesarean section, shortened gestation, management system, hereditary, hormonal, infectious disease and maternal immune system. Since there are many causative agents of retained placenta, the treatments which are commonly used for the retention of fetal membranes do not show the significant effects. Therefore, Basically, we have to avoid the occurrence of dystocia through genetic selection of dam and sire having minimal probability for RP, proper prepartum nutritional status and exercise. Special care should be paid for nutrition and vitamin supplementation, especially during the dry period.

It could be concluded that RP is an important problem which causes great economic losses and leave the animal subfertile even after treatment and recovery. So, it is recommended to control the condition rather than to treat it.

Keywords: Cows, Retention of placenta, Risk factors, Placental detachment, Dystocia.

INTRODUCTION

A fetal membrane is an essential organ for prenatal transfer of nutrients and oxygen from the dam to the fetus [1]. The other functions of placenta is it provides a reservoir of blood for the fetus, delivering blood to it in case of hypotension and vice versa, comparable to a capacitor [2]. When the fetus is born the placenta normally detaches within short time and is expelled. That is why it is referred to as the “afterbirth” [3]. The release of fetal membranes postpartum is a physiological process and involves loss of fetomaternal adherence, combined with contraction of uterine musculature and is usually accomplished within 6 hours of calving [4]. Normally fetal membranes drops within short time post partum (within 8 hrs of parturition), if it is retained up to 12 hrs then it is called as delayed removal and if retained for more than 24 hrs of parturition then it is called as ‘Retention of placenta’ (ROP). The key element in the pathogenesis of retained placenta in cattle is a failure of timely breakdown of the cotyledon-caruncle attachment after delivering the calf [5].

Retention of fetal membranes is the most common condition occurring in domestic animals following parturition [6]. Its incidence varies from 4.0-16.1%, but can be much higher in problem herds. However, it can be much higher in problem herds and also increases during summer with increased parity, milk yield in the previous seasons and following birth of male fetus [7,4]. Abortions, stillbirths and twin calvings resulted in increased incidence rates of 25.9, 16.4 and 43.8%, respectively. There are number of common causes that predispose for retention of fetal membranes, including mechanical, nutritional; managemental and infectious factors. Dystocia, caesarean section, uterine torsion, abortion, stillbirth, and twin birth are mechanical causes of RFM. While nutritional causes may be due to deficiency of protein, selenium, iodine, vitamin A and E and calcium deficiency during pregnancy. Managemental causes of retained placenta include stress hereditary, inbreeding and obesity [8]. Infectious causes is associated with brucellosis, salmonellosis, leptospirosis, and listeriosis [9, 10, 11]. Such retention creates a number of problems by allowing microorganisms to grow inside the uterus causing inflammation, fever, weight loss, decreased milk yield, longer calving intervals and may result in an open cow during the next year and if the infection is so bad the animal may actually die. Retained placenta is among the main reproductive disorders in farm animals especially, dairy cattle. It causes considerable economic losses in the herd due to decreased milk production, illness and treatment cost, beside a decreased market value of the animal [12, 13].

Diverse therapeutics has been employed for treatment of RFM. Manual removal, administration of intra-uterine and/or systemic antibiotics, injection of oxytocin, PGF_{2α} and β₂-receptor blockers; all assisted in prevention/treatment of RFM [14, 15, 16, 17]. Because RFM negatively affect milk production and cow's

fertility, effective treatment is crucial for improving puerperal performance of cows in order to raise their productiveness. Therefore, the aim of this paper is to review literatures on retention of fetal membranes in dairy cattle and to disseminate relevant information.

Literature review

Placenta

The placenta (Fetal membrane) is an organ that connects the developing fetus to the uterine wall to allow nutrient uptake, provide thermo-regulation to the fetus, waste elimination, and gas exchange via the mother's blood supply, fight against internal infection and produce hormones to support pregnancy. Placentas are a defining characteristic of placental mammals, but are also found in some non-mammals with varying levels of development [18]. It provides oxygen and nutrients to growing babies and removes waste products from the baby's blood. The placenta attaches to the wall of the uterus, and the baby's umbilical cord develops from the placenta. Other functions of placenta is it provides a reservoir of blood for the fetus, delivering blood to it in case of hypotension and vice versa, comparable to a capacitor [2]. The umbilical cord is what connects the mother and the baby. The essential materials pass to the developing fetus. When the fetus is born the placenta normally detaches within short time and is expelled. That is why it is referred to as the "afterbirth" [3]. The release of fetal membranes postpartum is a physiological process and involves loss of fetomaternal adherence, combined with contraction of uterine musculature and is usually accomplished within 6 hours of calving [4].

Retained placenta

Retention of placenta is the inability of fetal membrane to be expelled within 8 hrs after parturition and ranging of the retention from 8 to 48 hours post partum [19]

The normal physiological stages of birth during parturition include dilatation of parturient canal, delivery of the fetus and expulsion of the fetal membranes. In normal condition, fetal membranes are usually expelled within two to eight hours of parturition. Any retention of fetal membranes beyond 12 hours could be considered as pathological [20]. The incidence of retained placenta varies from 4-18% of calving [21, 22, 23]. The uterus normally contracts approximately fourteen times an hour immediately following parturition but the frequency gradually diminishes to once every hour at 42 hours. Delayed involution of the uterus is usually associated with retention of membranes. Retained placenta had a significant negative effect on milk yield for several weeks after calving [24, 25]. The interval from calving to first service and conception were higher in the retained placenta and increases the risk of fatty liver syndrome and ketosis [10]. Retained placenta delays the postpartum resumption of cyclic ovarian function and prolongs the interval from calving to first ovulation [26]. Early or induced parturition, dystocia, hormonal imbalances, and immune-suppression are risk factors in interrupting the normal process resulting in retention of the placenta. Systemic administration of antibiotics can be beneficial in treating metritis and collagenase injection enhances placenta release during fetal retention [19].

Mechanism or pathogenesis of placental retention(Recent knowledge on pathogenesis of placental retention in cattle [27]

The key element in the mechanisms of retained placenta in cattle is a failure of timely breakdown of the cotyledon-caruncle attachment after delivering the calf.

Mechanism of normal placental separation

Maternal immunological recognition of foetal MHC class I proteins expressed by trophoblast cells triggers an immune/inflammatory response that contributes to placental separation at parturition. The processes leading to normal separation and delivery of the placenta are multifactorial and begin before parturition (Fig 1). For example, it has been suggested that serotonin might also play a role in regulating bovine placental attachment [28]. High fetal and placental serotonin during pregnancy could help to maintain placental attachment by promoting placental cell proliferation [28] and inhibiting matrix metalloproteinase (MMP) activity [29]. Maturation of the fetal monamine oxidase enzyme system close to parturition results in the metabolization and subsequent decrease in serotonin, which in turn could promote placental separation and parturition [28].

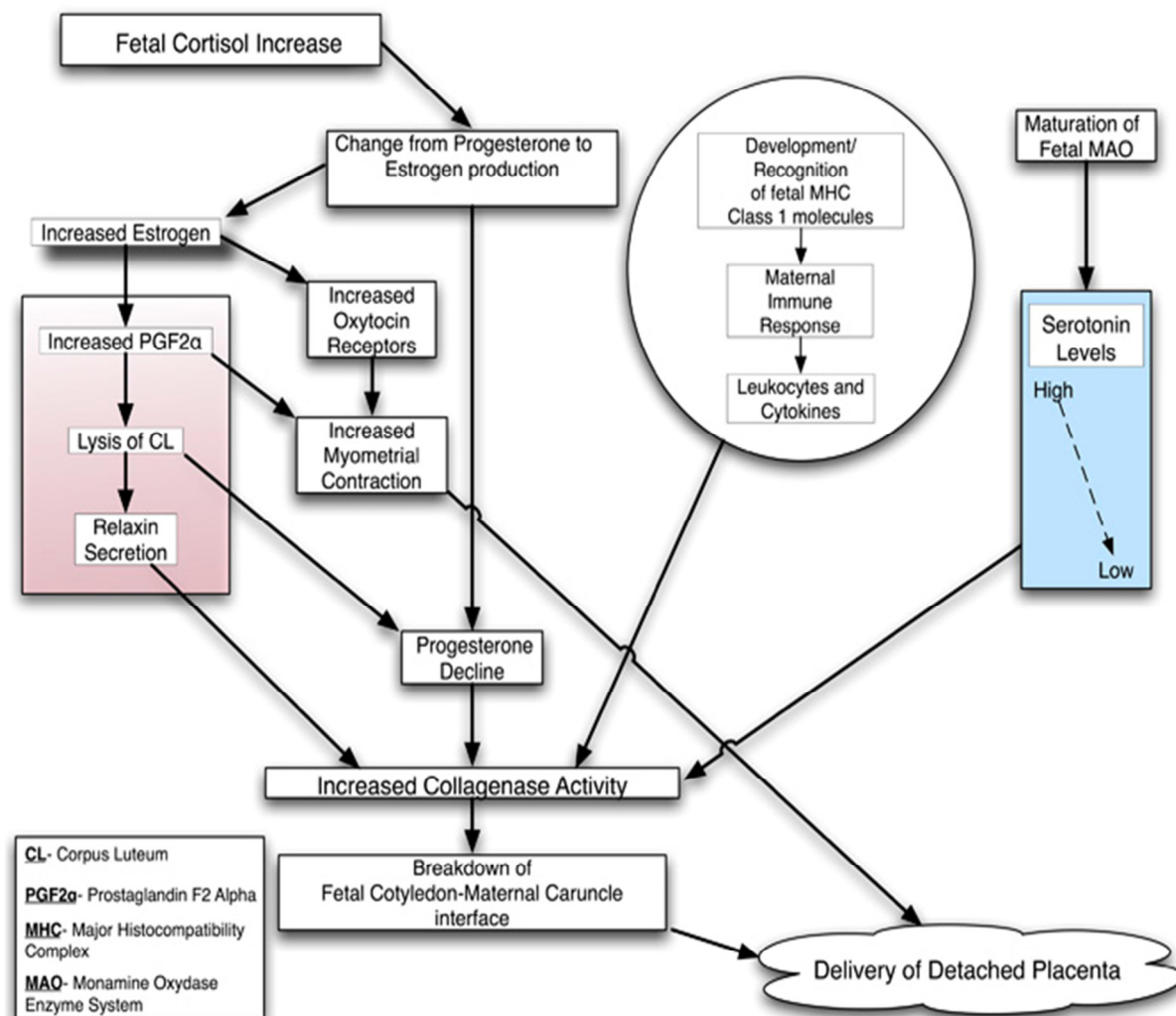


FIGURE- 1: Physiologic processes leading to the detachment of the placenta in dairy cows.

Mechanism of placental retention

Chemotactic factor for leukocytes is found in placentomes of cows with normal placental separation. It is absent in placentomes from cows with retained placentas. Blood leukocytes and neutrophils of cows with retained placenta are less reactive to chemotactic stimuli than in cows with normal placental separation. The cows with a greater degree of negative energy balance prepartum and higher non-esterified fatty acid (NEFA) concentrations were 80% more likely to suffer from retained placenta. Similarly these authors found higher risk for retained foetal membranes in cows with lower circulating vitamin E concentration. The recent data indicate that lack of uterine motility plays little or no role in the occurrence of retained placenta. Moreover cows with retained placenta have normal or increased uterine activity in the days after calving [15].

Etiology

ROP denotes the failure of the fetal villi to separate from the maternal crypts i.e. the lack of placental dehiscence. The causative factors are:

Infectious disease

Infectious disease causes of placental retention are behind the scope. Infectious diseases like Bovine Viral Diarrhea may cause RFP in cattle [30]. Brucellosis is a contagious bacterial disease of sexually mature animals and causing abortion and retained placenta [31, 32]. The disease is clinically characterized by abortion in the last trimester and retained placenta in the female whereas orchitis and epididymitis in male [33].

Managemental

Managemental causes of retained placenta include stress hereditary, inbreeding and obesity [8].

Lack of exercise and hypocalcemia are the most frequent causes of decreased myometrium contractility. Stress (Transportation, rough handling, poor feed conditions, Isolation from group, Lameness,) results in elevated corticosteroids and increased risk of placental retention.. Dairy producers have suggested that Poor health

management in herds can predispose animal to retention of placenta [34]. In addition to this deficiency of antioxidant, vitamin E and selenium may decrease chemo taxis and leukocyte numbers at the fetomaternal junction, thus contributing to the retention of fetal membranes [35]. Over-condition and under condition as well as managerial defects and environmental factors can result in retention of placenta [36].

Nutritional

Nutritional causes of RP are primarily due to the deficiency of feed during the last 6 to 8 weeks before calving specially when there is deficiency of content of minerals and vitamins in diet [37]. Heavy grain feeding may be associated with both higher milk production and increased risk of reproductive disorders such as dystocia, retained placenta, cystic ovaries, metritis other reproductive disorders [38]. Vitamin and mineral deficiency conditions such as selenium, vitamin E and vitamin A, B-carotene and disturbed Ca/P ratio can impair general immunity and may alter the competence of cellular self-defense mechanism and can increase the risk for placental retention and metritis [39]. High milking cows with a greater degree of negative energy balance prepartum and higher NEFA concentrations were more likely to suffer from RP [11]. On the other hand, over-conditioned cows were shown to be more sensitive to retained placenta and subsequent infertility than cows with normal body condition scores [40].

Cow's body weight

The percentage of retained placenta increases significantly with increasing live body weight of cows due to the increment in fat adipose tissues [41], which may result in trapping the steroid sex hormones.

Calves' birth weight

A significant increment of retained placental problem is happening with increasing fetal birth weight [41]. The reason could be due to pressure of the fetus on the placenta and fetal membrane [42], so that the attachment between the cotyledons and the fetal membrane become stronger these consequent in occurrence of placental retention.

Failure of maternal Immune Response

It is occurred due to failure of the maternal immune system to successfully degrade the placentomes at the end of pregnancy [15]. Maternal immunological recognition of fetal MHC class I proteins expressed by trophoblast cells triggers an immune inflammatory response that contributes to placental separation [43]. This lymphocytic activation was suppressed at the foeto maternal interface alongside the pregnancy course to avoid rejection of fetal allograft where the trophoblast secretes interferon-tau (IFN- τ) and both trophoblast and endometrium secrete prostaglandin E₂ and the endometrial glands secrete serpins (uterine milk proteins), all of which inhibit lymphocyte activation to keep on the embryo not rejected by the dam [44].

Hormonal

Placental separation occurs when fetal cortisol induces the production of the enzymes, 17 α -hydroxylase and aromatase in the placenta which favour oestrogen synthesis at the expense of progesterone synthesis [1]. Maternal plasma levels of oestradiol-17 β increase suddenly, while plasma levels of progesterone decline sharply immediately prior to parturition. It is supposed during the week before parturition, the level of estradiol reaches its maximum level to help the uterus to get rid of any remnant of fetal membranes. Therefore, a decreased level of estrogen may be indicated as a factor enhancing RP [45]. Spontaneous myometrial contractility is augmented by autocrine and paracrine release of PGF and parturition ensues. Disturbed endocrine function high progesterone and cortisol levels and low oestradiol level was traced in the blood cows with RP [46]. Increased progesterone level in RP may be due to failure of the placenta to produce specific steroidal enzymes that help in progesterone aromatization and its conversion to oestrogen [3].

Mechanical Causes of Retained Placenta

Difficult birth (calf too large for cow, backwards calf known as breech birth, one leg or head backwards), twins, late or premature birth, prenatal loss, induction of parturition with PGF₂, cesarean section and fetal monsters or emphysematous fetus (gas-filled fetus) are direct causes of dystocia and consequently to RP.

Failure of cotyledon-caruncle detaching mechanisms

The main cause of retained placenta is due to a lack of breakdown of the caruncle-cotyledon attachments after delivering the fetus [47]. The reasons could be due to infectious and/or noninfectious factors [48]. Primary attention has been often directed to infectious causes, but non infectious factors probably account for 70% or more of the cases [15]. Noninfectious causes are often multifactorial and are difficult to diagnose [49].

Risk factors

There are a number of risk factors associated with RFP, including Age, parity, induction of parturition by with PGF₂ α [50], Repeatability, stillbirth, shortened gestation [51], abortion of fetus [52], twinning [51], dystocia [24], fetotomy [53], Cesarean section [52], nutritional deficiencies such as vitamin E, selenium, and carotene [54], immunosuppression [55], metabolic disorders, especially milk fever.

Infectious agents such as bovine viral diarrhoea virus [30], brucellosis [31,56], leptospirosis, vibriosis, listeriosis, IBR, Trichomoniasis, *Listeria monocytogenes*, and etc. Whereas the exact mechanism responsible for

these factors is not completely known, the complex process of multiple hormones and biochemical events leads that disturbance in these events causes the RFM. The studies of physiological factors which are responsible for the detachment of fetal membranes help in the diagnosis of etiologies of RFM.

Reproductive Impact of Retained Placenta

The fertility of the dairy cows is affected when most cows in the herd suffer from retained placenta. These causes direct loss to the farmer due to delayed calving leading to a lengthy period between births (calving intervals) and hence low milk production. The adverse effects of RFM on reproductive performance of cattle are: delay in first service [57], reduction of pregnancy rate [58], increase in services per conception [59]. The RFM also leads to endometritis, puerperal metritis and mastitis [60] and these diseases ultimately cause the reduction in the fertility and milk production of cattle [55].

Mastitis

Although the main economic impact of ROP seems to be decreased milk production, more days open, decreased milk volume, milk from treated cows withheld), the correlation between ROP and mastitis is controversial. However, the economic losses as a result of mastitis could be due to reduced milk production, discarded milk, reduced cow sale value, drugs and veterinary services. It is unhygienic to milk a cow with decomposing afterbirth hanging on it [11].

Metritis

Retention of placenta and metritis are positively correlated. Cows with ROP had a significantly higher incidence of metritis than cows without ROP and also a significant difference was found between conception rates in cows with ROP and metritis [61]. Retention of placenta results from the presence of decomposing placental tissues, which provide a favorable environment for bacterial colonization. Coliform bacteria and high concentrations of endotoxins present in lochia of cows with ROP are potent inducers of prostaglandins and cytokines, favoring development of uterine infections [62]. Metritis result in decreased dry matter intake, and hence, multiparous cows with metritis in early lactation produce less milk than the healthy cows. This difference is greatest during the first 20 weeks of lactation [63].

Economic consequences of retained placenta

Retained placenta, one of the main causes of endometritis in cattle, causes economic loss [64, 65] Kossabati and Esslemont [66] calculated the direct cost of a case of retained placenta to be about £ 83, with an over-all cost of £ 298.29 (1995 prices).

Losses due to infertility and low milk production

In dairy cows retained placenta may be the cause of serious economic losses to the farmers as cows with retained placenta may develop bacterial infection and become ill and thus reduce production. Some may even die. Milk from cows with retained placenta is unfit for human consumption and therefore cannot be sold. The fertility of dairy cows is affected when most cows in the herd suffer from retained placenta. This causes a direct loss to the farmer due to delayed calving leading to a lengthy period between births (calving intervals) and hence low milk production. The retained fetal membrane causes considerable economic loss, especially when incidence exceeds the average of 5-10% [8]. The fertility of cows after retention of the placenta appeared to be affected. Generally, retention of placenta has great influence on productivity. For instance, retained placenta had a significant negative effect on milk yield for several weeks after calving and there is considerable milk loss as a result of difficult of calving [67].

Reduce conception rate

Varies studies reported that the conception rate of cows presenting retained placenta were significantly lower compared to normally calved cows [68]. The highest proportion of normal cows was conceived during the period from 61 to 90 days after parturition, while cows with retained placenta were conceived at more than 120 days after parturition [41].

Delaying post-delivery service interval

Placental retention is usually accompanied by delayed involution of the uterus [17], and adversely affects reproductive performance [69]. Cows with reproductive disorders had longer intervals from calving to first service and to conception and required more services per conception and lower pregnancy rate and conception to first service [68]. The period from parturition to the first service was longer in cows exhibiting retained placenta compared to normal ones [41].

Longer calving interval

Retention of placenta and metritis may cause prolonged calving interval and permanent infertility. Calving interval remained longer in cows revealing retained placenta as compared to normal cows [41, 69]. In general, the financial losses due to retained placenta in dairy cattle existed due to increased calving interval, increased culling rate, reduced conception rate, infertility, loss of milk production, the costs of veterinary service and drugs.

Treatment

It is important to clearly establish the objective of RP treatment. Many antimicrobial and hormonal treatments have been applied to cows with RP, generally without any reduction in risk of subsequent disease (such as displaced abomasum) or improvement in reproductive performance [17, 22, 70, 71, 72]. The use of antimicrobial therapy in the treatment of RFM has demonstrated conflicting results [14]. Postpartum metritis is common sequelae of RFM, and the rationale behind antibiotics for RFM is to prevent or treat metritis and its subsequent negative effects on fertility. Tetracycline antibiotics commonly used for intrauterine treatment in cattle, inhibit MMPs and might therefore interfere with the normal placental detachment mechanisms [73]. Daily intrauterine (IU) infusions of 5 g oxytetracycline for as long as the RP is in place reduced the incidence of fever from approximately 50% of cows with RP to approximately 30% of affected cows. However, the administration of intrauterine antibiotics did not reduce the incidence of metritis following RFM and could not improve fertility parameters [74].

And also ceftiofur (1.1 mg/kg IM q 24 h for 5 days) in cows with RP and fever was as effective (67% absence of fever by 10 days in milk (DIM)) as a combination of systemic and IU ampicillin and manual removal of the placenta; there was no difference in reproductive performance between the two treatments [75]. An experimental injection of collagenase into the placental end of the umbilical artery has been shown to facilitate separation of the placenta from the uterus [29]. Systemic antibiotics are believed to be beneficial in RFM cases [76].

The hormone which are used for treating retention of fetal membranes or retained of placenta are prostaglandins and oxytocin. These hormones play a role in uterine contraction and could be effective in treating RFM because of uterine atony. However, it is thought that uterine atony accounts for a very small percentage of retained placenta cases and numerous studies have not supported their use as a general treatment for RFM [77]. Oxytocin has long been advocated to expel the placenta after delivery. There are other advantages to the use of oxytocin after calving but it does not reduce the incidence of retained placenta. Oxytocin is already being secreted by normal cows at parturition and it helps contract the uterus and expel a placenta that is fully detached. The contraction of the uterus helps control bleeding from the various sites that may have been traumatized during delivery. If the placenta is not detached from the caruncles oxytocin will not hasten its passage [78]. Additionally oestradiol and the synthetic stilbene, stilboestrol, have been widely used in the treatment of retained placenta. They were thought to have beneficial effects by increasing uterine tone, and particularly by increasing the response to oxytocin [79].

Historically, manual removal of RP by manipulation and traction was practiced. There is no evidence that this practice produces beneficial results [80], and some evidence suggests that it is harmful [14]. Several studies indicate that approximately 50–80% of cows with untreated RP will have a temperature $>39.5^{\circ}\text{C}$ on at least 1 day within 10 days postpartum [57, 71, 75, 80, 81].

Manual removal can result in more frequent and severe uterine infections, when compared with more conservative treatment [14]. Bolinder et al., [14] found that manual removal prolonged the interval from calving to 1st functioning CL by 20 days. Additionally, intrauterine pathogenic bacteria were found in 100% of cows with manually removed RFM versus 37% of untreated cows at 3 weeks postpartum, and further 37% of treated versus 12% of untreated cows at 5 weeks postpartum. While current evidence does not support manual removal as an effective treatment for RFM, it is still commonly practiced [17]. And also as Drillich *et al.*, [72] demonstrated that 84.3% of the cows after manual removal of the placenta developed fever. Furthermore, remnants of cotyledons were found on caruncles, and hemorrhages and thrombi could be seen microscopically in cows after assumed removal of RFM [21]. Nevertheless, the manual removal of RFM together with the administration of tetracycline bolus is still the routine treatment [82, 83].

Prevention and control

The control of retained placenta needs to focus on the control of causative factors like abortions, premature calving, calving difficulties, and vitamin and mineral deficiencies. Milk fever and even sub-clinical calcium deficiency can be associated with an increased risk of RFM [84, 85] with older cows more at risk of lower blood calcium. Then it needs to be controlled. Good control of feeding and condition during the dry period and avoiding cows becoming overfat will also reduce the incidence of retained placenta. The herds with a history of selenium deficiency had a high incidence of RFM, and according to their suggestion supplementation of vitamin E and selenium can help to reduce placental retention [35, 86]. The synthetic form of vitamin E (alpha-tocopherol acetate) was found to be more effective than the natural form of vitamin E [35]. Supplementation with balanced vitamin and mineral mixture in prepartum period is considered a prophylactic step to avoid fetal membrane retention. Although these vitamins and minerals can be supplemented, correct pasture based diet formulation could prevent the need for additional supplementation. For example, Beeckman et al., [87] showed that grass clover silage and mixed silage were shown to have significantly more vitamin E than hay, maize or grain. The infectious diseases can be prevented by proper immunization against specific infection.

REFERENCE

- 1, Emtenan M Hanafi, 2011. Department of Animal Reproduction and AI, Veterinary Research Division, etiology of retained placental in dairy cattle. *Anim Prod Sci*, 14: 251-262.
- 2, Assad RS, Lee FY, Hanley FL, 2001. "Placental compliance during fetal extracorporeal circulation". *Journal of applied physiology (Bethesda, Md. : 1985)*. 90 (5): 1882–1886.
- 3, Ball P and A Peters, 2004. Reproduction in Cattle. Third Edition. Blackwell publishing. pp: 430
- 4, El-Malky, O, M Youssef, N Abdel-Aziz and A Abd El-Salaam, 2010. Postpartum Performance of cattles with retention of placenta.
- 5, Davies, R., Dalziel, R., Gibbens, J., Wilesmith, J., Ryan, J., Evans, S., Byrne, C., Paiba, G., Pascoe, S. and Teale, C., 2004. National survey for Salmonella in pigs, cattle and sheep at slaughter in Great Britain, 1999-2000. *J. Appli. Micro.*, 96: 750-760.
- 6, Noakes, D., Parkinson, J. and England, G., 2009. Veterinary Reproduction and Obstetrics. 9th Ed. China: Saunders Elsevier. Pp 418-425.
- 7, Ahmed WM, El-Ekhnawy KI, Dessouky HM, Zabal MM and Ahmed YF., 1999. Invisitations on retained fetal membranes in Friesian cows in Egypt. *Egyptian Journal of comparative Pathology*, 12: 160-177.
- 8, Joosten, I., M. F. Sanders and E. J. Hensen, 1991. "Involvement of major histocompatibility complex class I compatibility between dam and calf in the aetiology of bovine retained placenta." *Anim Genet* 22(6): 455-463.
- 9, Grohn, Y. T. and P. J. Rajala-Schultz, 2000. "Epidemiology of reproductive performance in dairy cows." *Anim Reprod Sci* 60-61:605-614.
- 10, Han, I. K. and I. H. Kim, 2005. "Risk factors for retained placenta and the effect of retained placenta on the occurrence of postpartum diseases and subsequent reproductive performance in dairy cows." *J Vet Sci* 6(1):53-59.
- 11, LeBlanc, S. J., T. F. Duffield, K. E. Leslie, K. G. Bateman, J. TenHag, J. S. Walton and W. H. Johnson, 2002. "The effect of prepartum injection of vitamin E on health in transition dairy cows." *J Dairy Sci* 85(6): 1416-1426.
- 12, Ahmed, W.M., H.H. El-khadrawy and A.R. Abel Hameed, 2006. Applied investigation on ovarian inactivity in buffalo heifers. In *Proc. of 3rd. Intl. Conf. Vet. Res. Div., NRC.*, pp: 1-15.
- 13, Maldonado, J. and S. Shreif, 2010. Livestock and Product Annual Global Agricultural International Network USDA Foreign Agricultural Service.
- 14, Bolinder, A., B. Seguin, H. Kindahl, D. Bouley and D. Otterby. 1988. "Retained fetal membranes in cows: Manual removal versus nonremoval and its effect on reproductive performance." *Theriogenology* 30(1):45-56.
- 15, Frazer, G. S., 2005. "A rational basis for therapy in the sick postpartum cow." *Vet Clin North America Food Anim. Pract.* 21(2):523-568.
- 16, Garcia, A., A. D. Barth and R. J. Mapletoft, 1992. "The effects of treatment with cloprostenol or dinoprost within one hour of induced parturition on the incidence of retained placenta in cattle." *Can Vet J* 33(3):175-183.
- 17, Peters, A. R. and R. A. Laven, 1996. "Treatment of bovine retained placenta and its effects." *Vet Rec* 139(22):535-539.
- 18, Pough et al. 1992. *Herpetology: Third Edition*. Pearson Prentice Hall:Pearson Education, Inc., 2002).
- 19, Beagley, J. C., Whitman, K. J., Baptiste, K. E. and Scherzer, J., 2010. DePhysiology and treatment of retained fetal membranes in cattle: A review.2; *Journal of Veterinary Internal Medicine*, 24:261–268.
- 20, Wetherill, G. D., 1965. Retained placenta in the bovine. A brief review. *Canadian Veterinary Journal*, 6: 290-294.
- 21, Paisley, L. G., Mickelson, W. D. and Anderson, P. B., 1986. Mechanisms and therapy for retained fetal membranes and uterine infections of cows: A review. *Theriogenology*, 25: 353- 381.
- 22, Eiler, H., 1997. Retained placenta. In: R.S Youngquist. (Editor), *Current therapy in large animal. Theriogenology*, W.B. Saunders, Philadelphia. Pp. 340-348.
- 23, Noakes, D. E., Parkinson, T. J. and England, G. C. W., 2001. Veterinary Reproduction and Obstetrics. Saunders, Pp. 383-472.
- 24, Rajala, P.J. and Grohn, Y.T., 1998. Effects of dystocia, retained placenta, and metritis on milk yield in dairy cows. *Journal of Dairy Science*, 81: 3172–3181.
- 25, Lucey, S., Rowlands, G.J. and Russell, A., 1986. Short-term associations between disease and milk yield of dairy cows. *Journal of Dairy Research*, 53: 7-15).
- 26, Opsomer, G., Grohn, Y.T., Hertl, J., Coryn, M., Deluyker, H., Kruif, A., 2000. Risk factors for post partum ovarian dysfunction in high producing dairy cows in Belgium: a field study. *Theriogenology*, 53: 841-857.
- 27, Davies CJ, Hill JR, Edwards JL, Schrick FN, Fisher PJ, Eldridge JA., 2004. Major histocompatibility antigen expression on the bovine placenta: Its relationship to abnormal pregnancies and retained placenta. *Anim. Reprod. Sci.* 82 – 83: 267 – 280.
- 28, Fecteau KA, Eiler H., 2001. Placenta detachment: Unexpected high concentrations of 5-hydroxytryptamine (serotonin) in fetal blood and its mitogenic effect on placental cells in the bovine. *Placenta*, 22:103–110.

- 29, Eiler H, Hopkins FM., 1993. Successful treatment of retained placenta with umbilical cord injections of collagenase in cows. *J Am Vet Med Assoc.*, 203:436-443.
- 30, Niskanen R, U Emanuelson, J Sundberg, B Larsson and S Alenius, 1995. Effects of infection with bovine virus diarrhoea virus on health and reproductive performance in 213 dairy herds in one country in Sweden. *Prev Vet Med*, 23: 229-237.
- 31, Aulakh HK, PK Patil, S Sharma, H Kumar, V Mahajan and KS Sandhu, 2008. A study on the epidemiology of bovine brucellosis in Punjab (India) using milk-ELISA. *Acta Vet Brno*, 77: 393-399.
- 32, Kebede, FU, S Ibrahim, I Ajogi and BJ Olaniyi, 2011. Prevalence of retention of placenta and risk factors assessment in cattle herds in Jigawa state. *ISRN Vet Sci* 10.5402/2011/132897.
- 33, Radostits O, Blood D and Gay C., 1994. *Veterinary Placenta*, 27: 794-798. pp: 999- 1002 pregnant and lactating Egyptian cattles. 1st Sci. Conf. Vet. Physio., Beni-Suef., pp: 77-82.
- 34, Fricke PM., 2001. Review: twinning in dairy cattle. *Prof Anim Sci*, 17: 61-67.
- 35, Bourne N, R Laven and DC Wathes, 2007. A meta-analysis of the effects of vitamin E supplementation on the incidence of retained fetal membranes in dairy cows. *Theriogenology*, 67: 494-501. *Bovine Ann Med Vet*, 143: 91-118.
- 36, Hayirli A, RR Grummer, EV Nordheim, and PM Crump, 2002. Animal and dietary factors affecting feed intake during the prefresh transition period in Holsteins. *Dairy Sci*, 85: 3430-3443.
- 37, Spears, J.W. / Weiss, W.P., 2008. Role of antioxidants and trace elements in health and immunity of transition dairy cows. *The Veterinary Journal*, 176(2008)1, 70-76.
- 38, Grimard B, S Freret, A Chevallier, A Pinto, C Ponsart, and P Humblot, 2006. Genetic and environmental factors influencing first service conception rate and Jorritsma R, T Wensing, TAM Kruip, PL Vos, and JPTM Noordhuizen 2003: Metabolic changes in early lactation and impaired reproductive performance in dairy cows. *Vet Res* 34: 11-26 *J Agric Sci Mansoura Univ*, 30: 6532.
- 39, Ahmed WM and MM Zaabal, 2009. Analyzing the immunogenetic constituents of dams, sires and calves in relation to placental retention in a Frisian herd. *Global Vet*, 3: 32-36.
- 40, Badinand F and G Sensenbrenner, 1984. Non-delivrance chez la vache: doiuiées nouvelles a propos d'une enquete epidemiologique. *Point Vet*, 16: 483-496.
- 41, Gaafar, Sh A-Shamiah and MA Sh M-Abu El-Hamd, 2010. Factors related to the incidence of retained placenta in Friesian cows kept under Egyptian conditions.
- 42, Deyab, 2000. The effect of different treatment upon the postpartum involution of uterus and retention of the placenta in dairy cows. *The Veterinary Medical Review*, 1977, 1: 36-43.
- 43, De-Mouzon, H and M Millo, 2006. The placenta cytokine network and infammatory signals.
- 44, Hansen, P and W Liu, 1996. Immunological aspects of pregnancy: concepts and speculations using the sheep as a model. *Anim Reprod Sci*, 42: 483-493.
- 45, El-Nemer I, N Hazza and S Emara, 2000. Changes in thyroid and sex hormones in serum of cattle with retention of placenta.
- 46, Michal, K, M Edward and M Hanna, 2006. Some hormonal and biochemical blood indices in cows with retained placenta and puerperal metritis. *Bull Vet Inst Pulawy*, 50: 89-92.
- 47, Martin, G. J., 1996. *Ethnobotany; People and Plant Conservation Manuals*. Worldwide Fund for Nature International, Chapman and Hall, London | *Merck Veterinary Manual* (1986). Merck and Co., Inc. Rahway, NJ., USA. P682 683. 682-683 *Med* 1999;41:1-35. *medicine*, 8th Philadelphia: W.B. Saunders. pp: 1177-1190.
- 48, Moizur Rahman M, M Zahan, A Kader Md and KM Mozaffor Hossain, 2013 Retained placenta of dairy cows associated with managemental factors in Rajshahi, Bangladesh, *Vet World*, 64: 180-184, doi: 10.5455/vetworld.2013.180-184.
- 49, Hanzen C, Drion O, Lourtie, Depierreux C and Christians E., 1999. La mortalite embryonnaire. Aspect cliniques et facteur setiologiques dans l' espece bovine. *Annales de MédecineVétérinaire*, 143: 91-118.
- 50, Terblanche HM, Kritzing LJ, Van Heerden JS, 1976. Induced parturition in cattle. 1. Clinical studies. *J S Afr Vet Assoc.*, 47:113-115.
- 51, Muller LD, Owens MJ., 1974. Factors associated with the incidence of retained placentas. *J Dairy Sci*. 57: 725-728.
- 52, Joosten I, Van Eldik P, Elving L, Vander Mey GJW., 1987. Factors related to the etiology of retained placenta in dairy cattle. *Anim. Reprod. Sci*. 14: 251 - 262.
- 53, Wehrend AT, Reinle K, Herfen H, et al., 2002. Fetotomy in cattle with special reference to postoperative complications-an evaluation of 131 cases. *Dtsch Tierarztl Wochenschr*, 109:56-61.
- 54, Julien WE, Conrad HR., 1976. Selenium and vitamin E and incidence of retained placenta in parturient dairy cows. *J. Dairy. Sci*. 59: 1954 - 1959.
- 55, Laven RA, Peters AR., 1996. Bovine retained placenta: Aetiology, pathogenesis, and economic loss. *Vet. Rec*. 139: 465 - 471.
- 56, Kebede, T., Ejeta, G., Ameni, G., 2008. Seroprevalence of bovine brucellosis in smallholder farms in central

- Ethiopia (Wuchale –Jida district). *Rev. Méd. Vét.* 15 (9), 3 – 9.
- 57, Stevens RD, Dinsmore RP., 1997. Treatment of dairy cows at parturition with prostaglandin F2 a or oxytocin for prevention of retained fetal membranes. *J. Am. Vet. Med. Assoc.* 21: 1280 – 1284.
- 58, McDougall S., 2001. Effects of periparturient diseases and conditions on the reproductive performance of New Zealand dairy cows. *NZ. Vet. J.* 49: 60 – 68.
- 59, Holt LC, Whittier WD, Gwazdauskas FC., 1989. Early postpartum reproductive profiles in Holstein cows with retained placenta and uterine discharges. *J. Dairy. Sci.* 72: 533 – 539.
- 60, Bruun J, Ersb AK, Alban L., 2002. Risk factors for metritis in Danish dairy cows. *Prev. Vet. Med.* 54: 179 – 190.
- 61, Youngquist RS and WR Threlfall, 2007. *Current Therapy in large Animal Theriogenology*, 2nd ed., Saunders, and imprint of Elsevier Inc, USA.
- 62, Dohmen, 2005. Effect of retained placenta on postpartum reproduction performance of Frisian cows. *Egypt J Anim Prod*, 40: 111-121.
- 63, Wittrock JM, KL Proucifoot, DM Weary, and MAG von Keyserlingk, 2011. Short communication: metritis affects milk production and cup rate of Hosten multiparous cows differently. *J Dairy Sci*, 94: 2408-2412. <http://dx.doi.org/10.3168/jds.2010-3697> PMID: 21524531.
- 64, Joosten I, Stelwagen J, Dijkhuizen AA., 1988. Economic and reproductive consequences of retained placenta in dairy cattle. *Vet Rec.*, 123, 53-57.
- 65, Kaneene JB, Miller R., 1995. Risk factors for metritis in Michigan dairy cattle using herd- and cow-based modelling approaches. *Prev Vet Med.*, 23, 183-200.
- 66, Kossaibati MA, Esslemont RJ., 1997. The costs of production diseases in dairy herds in England. *Vet J.*, 154, 41-51.
- 67, Dematawewa CMB, and PJ Berger, 1997. Effect of dystocia on yield, fertility, and cow losses and an economic evaluation of dystocia scores in Holsteins. *J Dairy Sci*, 80: 754-761.
- 68, Shiferaw Y, BA Tenhagen, M Bekana, and T Kassa, 2005. Reproductive disorders of crossbred dairy cows in the central highlands of Ethiopia and their effect on reproductive performance. *Trop Anim Health Prod*, 37: 427-441.
- 69, Swiefy AS., 2003. Effect of retained placenta on postpartum reproduction performance of Frisian cows. *Egypt J Anim Prod*, 40: 111-121.
- 70, Gilbert, R.O., Schwark, W.S., 1992. Pharmacologic considerations in the management of peripartum conditions in the cow. *Veterinary Clinics of North America: Food Animal Practice* 8, 29–56.
- 71, Stevens, R.D., Dinsmore, R.P., Cattell, M.B., 1995. Evaluation of the use of intrauterine infusions of oxytetracycline, subcutaneous injections of fenprostalene, or a combination of both, for the treatment of retained fetal membranes in dairy cows. *Journal of the American Veterinary Medical Association* 207, 1612–1615.
- 72, Drillich, M., Mahlstedt, M., Reichert, U., Tenhagen, B.A., Heuwieser, W., 2006a. Strategies to improve the therapy of retained fetal membranes in dairy cows. *Journal of Dairy Science* 89, 627–635.
- 73, Eiler, H., Hopkins, F.M., 1992. Bovine retained placenta: effects of collagenase and hyaluronidase on detachment of placenta. *Biology Reprod.*, 46: 580-585.
- 74, Laven, R.A. and Peters, A.R., 1996. Bovine Retained Placenta: Aetiology, Pathogenesis and Economic Loss. *Veterinary Record*, 139, 465-471.
- 75, Drillich, M., Sabin, M., Sabin, H.-J., Heuwieser, W., 2003. Comparison of two protocols for the treatment of retained fetal membranes in dairy cattle. *Theriogenology* 59, 951–960.
- 76, Risco CA, Hernandez J., 2003. Comparison of ceftiofur hydrochloride and estradiol cypionate for metritis prevention in dairy cows affected with retained fetal membranes. *Theriogenology*, 60: 47-58.
- 77, Drillich M, Schroder A, Tenhagen BA., 2005. Efficacy of a treatment of retained placenta in dairy cows with prostaglandin F2a in addition to a local antibiotic treatment. *Dtsch Tierärztl Wochenschr*, 112: 174-179.
- 78, Miller, B.J. and J.R. Lodge. 1984. Postpartum oxytocin treatment for prevention of retained placentas. *Theriogenology*. 22:385-388.
- 79, Roberts SJ., 1986 *Veterinary Obstetrics and Genital Diseases*. Wood stock, V.T. and Roberts, S.J., 373-393.
- 80, Drillich, M., Reichert, U., Mahlstedt, M., Heuwieser, W., 2006b. Comparison of two strategies for systemic antibiotic treatment of dairy cows with retained fetal membranes: preventive vs. selective treatment. *Journal of Dairy Science* 89, 1502–1508.
- 81, Callahan, C.J., Horstman, L.A., 1987. Treatment of early postpartum metritis in a dairy herd: response and subsequent fertility. *Bovine Practitioner* 22, 124–128.
- 82, Hehenberger, E.M., Doherr, M.G., Bodmer, M., Steiner, A. and Hirsbrunner, G., 2015. Diagnose und Therapie von Nachgeburtverhalten, puerperaler Metritis und klinischer Endometritis beim Rind: Ergebnisse der Online-Umfrage bei Schweizer Tierärzten. I Nachgeburtverhalten. *Schweiz Archiv Tierheilk*, Accepted for Publication.
- 83, Sheldon, I.M., Lewis, G.S., LeBlanc, S. and Gilbert, R.O., 2006. Defining Postpartum Uterine Disease in

Cattle. *Theriogenology*, 65, 1516-1530.

84, Zhang, W.-C., Nakao, T., Kida, K., Moriyoshi, M., & Nakada, K., 2002. Effect of Nutrition during Pregnancy on Calf Birth Weights and Viability and Fetal Membrane Expulsion in Dairy Cattle. *Journal of Reproduction and Development*, 48(4), 415–415.

85, Melendez, P., Donovan, G. A., Risco, C. A., & Goff, J. P., 2004. Plasma mineral and energy metabolite concentrations in dairy cows fed an anionic prepartum diet that did or did not have retained fetal membranes after parturition. *American Journal of Veterinary Research*, 65(8), 1071–1076.

86, Allison RD and RA Laven, 2000. Effect of vitamin E supplementation on the health and fertility of dairy cows: a review. *Vet Record*, 703-708.

87, Beeckman, A., Vicca, J., Van Ranst, G., Janssens, G. P. J., & Fievez, V., 2010. Monitoring of vitamin E status of dry, early and mid-late lactating organic dairy cows fed conserved roughages during the indoor period and factors influencing forage vitamin E levels: Vitamin E content of forage in organic dairy farming. *Journal of Animal Physiology and Animal Nutrition*, 94(6), 736–746.